



NANOTECHNOLOGY TARGETING DRUG DELIVERY WITH BIOSENSOR - A REVIEW

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Abstract

In the development of drug delivery technologies, a group of nanotechnology techniques are being used. The advancement of the use of nanotechnology in medicine, known as nanomedicine, to create novel therapeutic and diagnostic approaches through the use of meticulously engineered materials on this long scale. In terms of dimension, form, and materials, the nanoparticles used in drug delivery have several styles. Particles and medication stability, in terms of drug loading capacity, the characteristics of each particle vary depending on the type of substance, and the classifications of the substances can often differ, such as nanospheres, nanocapsules, micelles, dendrimers, nanotubes and fullerenes, polymers with nanoparticulate drug targeting mechanisms and biosensory targeting mechanisms.

Keywords: Nanotechnology, drug delivery, dendrimers, electrochemical sensor.

Introduction

A big issue around the globe is seeking new and groundbreaking cancer treatments[1]. The therapeutic effectiveness of certain malignant tumors has increased dramatically with the rise of alternatives to cancer care and the idea of individualized therapy. Chemotherapy is a traditional way of treating cancer and is commonly used. Although chemotherapy operates through a variety of different mechanisms, its key role involves destroying vigorously developing cells indiscriminately, Tumors and normal cells that cause severe side effects, such as bone marrow depletion, hair loss, and gastrointestinal reactions, including [2]. The creation of drugs that target tumor cells more accurately. The subject of a significant cancer-related proportion studies over few past decades has thus been, rather than normal cells. While the introduction of personalized treatment has made huge advances in precision therapy, certain inevitable side effects are still present, And there has always been anxiety over the rise of opioid resistance. Cancer is death cause of a second leading at present, and for many tumors, therapies current are unsuccessful. Therefore, more and more studies are finding accurate cancer treatment and drug resistance solutions. Nanotechnology has been widely used in medicine over the last few decades, Including applications for detection and care and targeting tumors in a better and more effective way. Many advantages in cancer treatment have been demonstrated by nanoparticle based system of drug delivery, like pharmacokinetics, tumor cell precise targeting, reduction of health risks, and drug resistance [3,4]. NPs used in system of drug delivery are typically engineered depending Based on their size and features, depending on the neoplasm's pathophysiology. Mechanically, cancer therapy NP-carriers attack cells of tumor per NPs carrier effect and positioning effect after ingestion of



targeting content. Next, in order to cause killing, they drugs release into cells of tumor. Common chemotherapy agents and nucleic acids contain medications found within the nano-carriers, proposed that they may play a major role gene therapy and cytotoxic [5]. In addition, NPs have a forum for such poorly soluble medications that can both encapsulate and circulate drugs[6,7]. Nanotransmitters can increase the half-life of drugs and induce aggregation in tumor tissues given the size and surface characteristics of NPs and their role in improving permeability and retention[8, 9]. However, The targeting mechanism defends normal cells from medication cytotoxicity, helping to alleviate the side effects of chemotherapy treatment. [10]. Furthermore, paclitaxel-bound nanomaterials albumin reported less adverse effects and accepted doses were greater than solvent-based taxanes[11]. In addition to gene therapy and chemotherapy, several trials have reported the use of NP medicines in chemotherapy and ablation therapy for cancer [12,13]. It is suspected that the nanoparticle-based drug delivery method can boost Immunotherapy and reversion of the tumor's immunosuppressive microenvironment [14]. An rising number of nanotherapeutic medications have been launched or have achieved commercial stages in previous seasons. A randomized trial in the first phase was conducted in 2010 to administer small interfering RNA (siRNA) to solid cancer patients using a targeted nanoparticle-based platform[15]. Efficacy of an effectively targeted chemotherapeutic docetaxel-containing polymeric nanoparticle for more advantageous tumor therapy has been documented in another clinical trial relative to the solvent-based DTXL formulation [16]. NP-based drug delivery mechanisms in the arena of, the development of NPs hybrid has seen much more advances. In order to increase the function and stability of each system of drug delivery, the properties of multiple NPs are integrated into hybrid NPs. [17]. In addition, when anti-tumor multidrug resistance (MDR) is involved,, NPs have demonstrated some benefits they offer Pharmacological inhibition of the function of certain drug resistance operations, such as flow vectors on cell membranes, is exacerbated by treatment platforms[18]. NP- therapy based has already been shown to have promise in many forms of cancers , including breast cancer, to resolve MDR[19], Ovarian cancer [20], and cancer of the prostate [21]. Medicine nanotechnology has opened up a new level of cancer therapy and more in-depth research is needed to synthesize these two areas. This analysis examines the underlying rules of applying the methodology of the nano-carrier to the treatment of cancer, addresses the emerging issues and discusses future avenues for study.

Nanoparticles of Medical Uses

according to size, materials and shape nanoparticles for drug delivery provide various architectural designs. The features in terms of drug potential loading, and drug fixation particle, drug release speeds, and target delivery efficiency, every particle differs.. To study the various NP systems is beyond the reach of this discussion. A partial summary, however, displays the variety available.

Dendrimers

Regularly branched with a multifunctional central core atom, there are three-dimensional treelike structures. By polymerization, the branch units can originate from or be synthesized from the central



nucleus from the peripheral and terminate at the center molecule. Steric division length limits, and with limited molecular size but large molecular weights, dendrimer types are spherical shaped. Drug molecules can be bound on surface dendrimer to functional groups or sheltered in channels of dendritic of the sphere 's internal environment[22,23]. A number of carrier molecules, both hydrophobic and hydrophilic, can host these nanoparticles and drugs, effective transfer agents are useful. (Fig. 1).

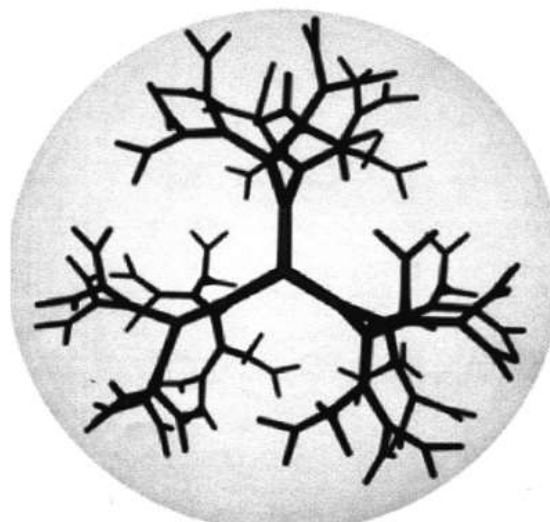


Fig. 1. Dendrimers. [23].

i. Dendrimers as Carriers of Drug Distribution

Due to their physicochemical structure, as drug delivery agents, dendrimers have also been studied extensively within Internal gap zones and structural classes of the exterior or periphery layer. In addition, the controllable shape, scale, Due to the sequential nature of the most typical synthetic procedures, inner and final functional groups, permeability, hydrodynamic length, molecular weight, molecular geometry, and variation of the internal and external charge can be constructed and incorporated into the final dendrimer structure [25-32]. Dendrimers have been used to protect all these properties of drugs, hormones, antibodies and other bioactive substances (Fig 2) [32-40].

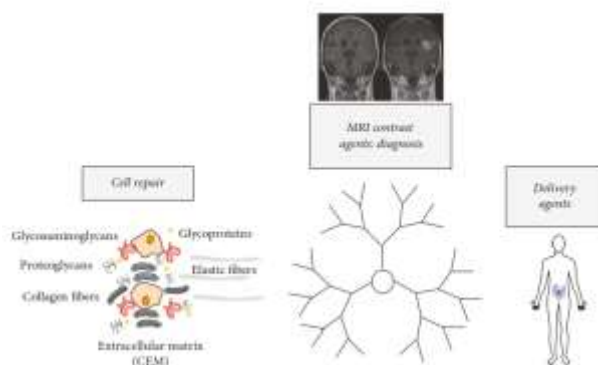


Fig 2: Schematic depiction of the use of dendrimers in various medical applications: repair of cells, diagnosis and delivery of drugs [40].



ii. Modified Electrodes with Dendrimers

There is a rising need for effective, cost-competitive, In medicinal chemistry, the food industry and various environmental industries, electrochemical sensors are precise and reliable. The most famous technique in this regard is the surface modification of conductor electrodes acting as supportive substances; in particular, the layer-by-layer (LBL) module method is among the most effective methods for the design and expansion of electrochemical sensors who have used chemical or biological protein molecules [41–43], viruses [44], DNA [45], nanoparticles [46], and dendrimers [47–79] Specific properties [47–50]. Their peculiar supramolecular properties have been used in the special case of dendrimers to identify and measure molecules of biological significance in LBL electrochemical sensors. Use of fundamental physicochemical characteristics, such as the capacity to encapsulate, Molecular protectionism, hydrophobicity/hydrophilicity biocompatibility, regulated dispersion and electrons kinetic influence [51-54], dendrimers are actually located as a popular characteristic of chemical surface substances for various electrochemical sensor manufacture.[55–58].

Micelles

The spherical or globular configurations of micelles are shaped in a liquid environment as hydrophobic side component molecules surrounding the main core of the sphere. Then the molecules' hydrophilic ends are liquid contact with the atmosphere covering the structure of the micelle and Compose a mantle. Micelles are useful for transporting water-insoluble drugs to the hydrophobic cardiac heart[23].

Nanospheres

These nanostructures are spherical constructions with a matrix composed of network in which the compound is dispersed through trapping and binding, the spherical forms of these nanoparticles consist of a matrix organization in which the substance is dispersed by trapping, binding[59] (Fig. 3).

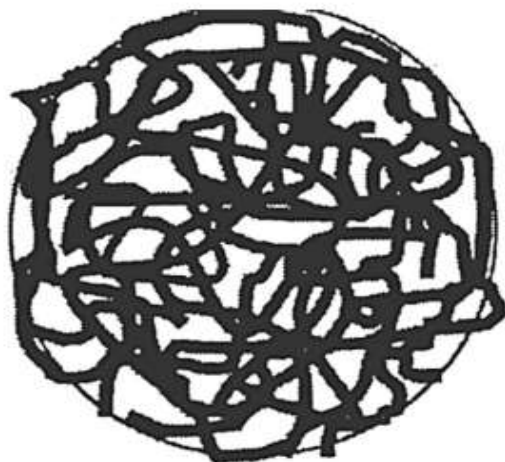


Fig.3. Nanospheres. (Reprinted with permission [59])



Nano-Capsules

These particles are vesicular structures with even a center or heart area to which a medication is limited. A polymeric membrane covers the heart of the outer shell to which it is possible to bind surface attached targeting ligands or antibodies. Solids, liquids, or gas may be the main material, and the main atmosphere may both oily or aqueous [59] (Fig. 4).

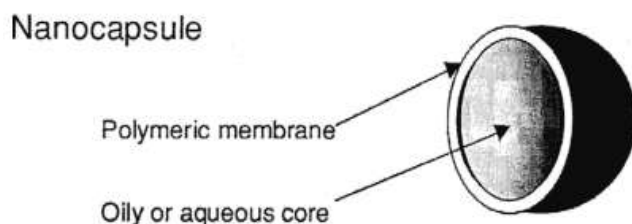


Fig. 4. Nanocapsules. (Reprinted with permission [59])

Nanotubes and Fullerenes

Such particles are a group of carbon molecules in the form of an ellipsoid tube or hollow sphere. Because of their resemblance to buckminster Fuller's geodesic dome form, Bucky spheres "bucky spheres." Spherical fullerenes are 60 bucky-tubes that are single or multi-walled with a hollow cage like structure. Within fullerenes and tubes and antibodies or ligands, atoms may be stuck to the surface for attack. [60] (Fig. 5).

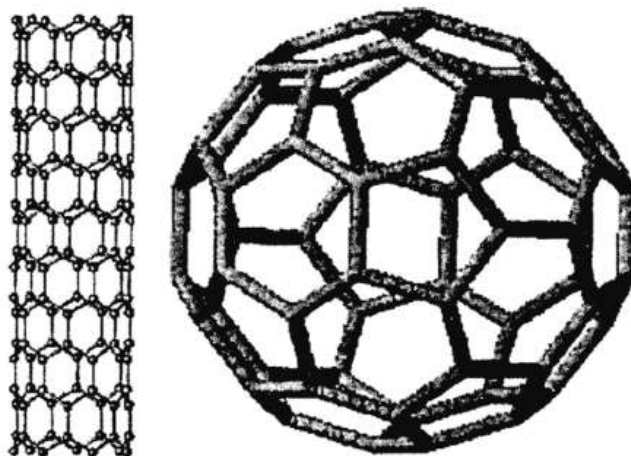


Fig. 5. Fullerenes and nanotubes. (Reprinted with permission [60]).

Liposomes

Liposomes are composed of an aqueous nucleus surrounded by a single nucleus or more phospholipid and cholesterol layers that make up a bilayer of lipids. Liposomes can prepare and hold hydrophilic factors in the aqueous environment and hydrophilicity substances in the lipid space due to this specialized structure. [61]. Liposomes are more biocompatible than other synthetic compounds, since their structure is close to that of the cell membrane. Moreover, different surface modifications with ligands and changes charge make it useful for the delivery of specific drugs tasks to coat liposomes



with polyethylene glycol(PEG). Liposomes have many extra advantages, like nanomedicines or systems for drug delivery. Liposomes protect the loaded medicine from decaying and preventing repeated exposure to the condition of the drug, which may delay the rate of drug release [62-64]. The lipid bilayer is stabilised by lipid species, such as cholesterol and constrictive saturated lipids, to avoid serum protein attacks and to reduce drug degradation. [62,63]. How to monitor their delivery and elimination in vivo, however, is the current problem facing the production of liposomes as drug carriers.

Polymers

It is possible to define polymers as: polymeric materials like proteins, glycans, peptides, cellulose and starches, such as polylactic acid (PLA) and polylactic acid (PLGA) synthesized from natural monomers such as polyhydroxybutyrates[65]. Types of synthetic polymers, including dendrimers, liposomes and micelles, are a diversification mode for fabricating a range of nanostructures (Fig. 6).

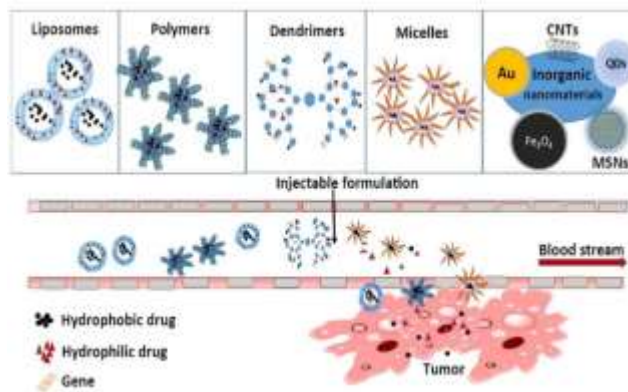


Fig 6. Nanomaterials that are used as carriers of drugs cancer therapy [65].

Challenges Posed By the Use of Nanocarriers to Extend Patient Survival

There are some biological properties of some solid tumors distinct based on those of normal tissues [66]. Abnormal properties of tumors, including biological influenced vascular system, irregular extracellular matrix and excessive interstitial fluid pressure, can produce limitations that undermine the efficacy of nanotherapeutic delivery [66,67]. There are also extravascular obstacles that need to be resolved, in which nanoparticles can extravasate but are unable to penetrate tumor ECM[68]. With its elevated blood pressure and decreased clearance of the venous and lymphatic, it is well understood that, High interstitial fluid pressure is caused by the irregularity of the tumor vasculature, resulting in nutrient diffusion and chemotherapy very inefficient in the tumor, thereby posing challenges for the successful diffusion of nanocarriers [69]. Due to their biocompatibility, biodegradability, and mechanical characteristics, The most commonly used biodegradable nanocarriers are liposomes and polymers. However, these nanocarriers can only give a fleeting extension of patient survival due to adverse effects and still-unclear association pathways between nanoparticles, the tumor microenvironment, and tumor cells.



Cancer Treatment NPs

Usually, There are specific sizes, shapes and surface characteristics of the NPs used in medical care, as these three characteristics have a significant effect on optimizing nano-drug distribution and thereby controlling clinical performance [70]. NPs with a maximum of 10 to 100 nm in diameter are commonly considered ideal for cancer therapy because then they can deliver the drug effectively and generate an improved gain of conductivity and retention (EPR). Smaller molecules can effectively remove into human cells (below 1-2 nm) from normal vascular system and can easily be filtered (below 10 nm in diameter) into the kidneys[71]. It is possible that particles larger than 100 nm are removed from circulation by phagocytes [72]. Moreover, their bioavailability and half-life can be influenced by the surface characteristics of NPs[73]. NPs are thus usually modified to be hydrophilic, raising the circulation drug time span and expanding their tumor penetration and aggregation[74;75]. Collectively, in cancer treatment, the different features of NPs decide their therapeutic effect.

Targeting Mechanisms

Specifically, cancer cell targeting is a crucial function since it improves therapeutic efficacy while shielding normal cells from cytotoxic effects, nano-carriers for drug delivery. To investigate the targeting nature of NP-based drugs, various experiments have been carried out. It is important to know cancer genetics and the relationship with nano-carriers and cancer cells in first order to further resolve the problems of cancer detection and nano-carrier device architecture. It is usually possible to separate the targeting processes In two classifications: active targeting and passive targeting (Fig 7).

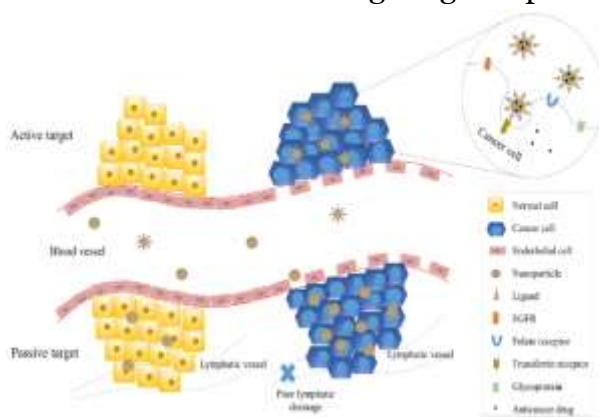


FIG 7 | Passive and aggressive labeling of cancer cells against NPs. Targeting NPs increases treatment efficiency and reduces systemic toxicity [76].

Targeting Cells for Cancer

Transferrin, a category of serum glycoprotein, has a role in carrying iron to cells. In too many solid cancerous cells, the transferrin mechanisms are over-expressed and are active in healthy cells at low concentrations. Transferrin-conjugated NPs are therefore used to distribute cancer therapy drugs as an effective targeting system[76-79]. NPs Transferrin-modified have been shown to demonstrate better



efficiency of cellular absorption and improved delivery of intracellular drugs compared to unmodified NPs [80,81]. In addition, evidence reveals that transferrin-conjugated polymer NPs play a significant role in the growth of drug-resistant therapies[82].

Mechanisms of Nps in the Overcoming of Drug Resistance

Drug tolerance is also a significant issue in cancer research, provided that cancer treatment methods are rising. Resistance to multidrugs refers to refusing various forms of cancer therapy, Lead to growth of tumor and bad prognosis. In tumor drug resistance pathways, faulty apoptosis - inducing machinery, interstitial pressure gradient and acid and hypoxic cancer micro - environment are concerned, such as cellular and biochemical influences Such as overexpression (e.g. efflux transporter) of ATP binding cassette (ABC) transporters[83]. It has been shown that nanotechnology for drug delivery for cancer care plays an significant role in resolving drug resistance.

Targeted Nanoparticles Delivery

Optimally, in general for chemotherapeutic agents to function in treatment for cancer, following transmission, they should still be able to reach the desired tumor tissues by breaching barriers in the system with minimal loss of volume or flowing blood activity. Second, drugs must be able to precisely kill cancerous cells without killing normal cells after entering the tumor tissue through a controlled release process of the active type. Both of these two important approaches was linked to enhancements in the survival and quality of care for patients by efficiently raising the rate of intracellular opioids and growing dose-limiting toxicity. More and more, for successful drug carrier structures, nanoparticles seem to have the ability to meet all of these criteria.

Scale and Characteristics of the Surface of Nanoparticles

Nanoparticles should be able to remain for a considerable period of time in the circulation without anyone being removed in order to successfully administer medicine to the intended tumor tissue. Based on their size and surface properties, The reticuloendothelial system, such as the liver and spleen, is normally stuck in circulation by conventional unmodified surface nanoparticles[84]. By altering their size and surface features, it is possible to monitor the fate of injected nanoparticles. Oh. Uh. Uh. Uh. Size. One of the nanoparticles' advantages is that they can be tailored to their dimension. In order to avoid their accelerated leakage into blood capillaries, the liver and spleen should have been the size of the nanoparticles used in a targeted drug delivery., for beginners, are large enough but small enough to evade suspicion by fixed tumor cells in the reticuloendothelial. In the spleen and fenestra of the Kuffer liver cells, the size of the sinusoid ranges from 150 to 200 nm[85] and the size of the leaky tumor vasculature distance between endothelial cells can differ from 100 to 600 nm[86]. Consequently, to enter tumor tissues by going through these two specific vascular systems, Nano-particles can be up to 100 nm in size. Features of surfaces. The surface characteristics of nanoparticles, in addition to their size, in relation to their macrophage capture, These are indeed an essential factor in deciding their cycle time and destiny throughout circulation. Avoiding detection of macrophages, nanoparticles should



preferably have a hydrophilic surface[87]. This can be achieved in two ways: by repelling plasma proteins from opsonization, the surface coating of nanoparticles with a hydrophilic polymer such as PEG protects them; alternatively [88].

Nanoparticles Potential to Resolve Resistance of Drug

Drug resistance has arisen to reduce the therapeutic potency of chemotherapeutic agents as a significant hurdle. Among multiple drug resistance pathways, the best known and most thoroughly studied P-glycoprotein is[89]. It has been proposed that nanoparticles would be able to sidestep resistance caused by P-glycoprotein. One possible mechanism is that when reaching the cell through being enveloped in an endosome, nanoparticles can avoid detection by the P-glycoprotein efflux pump, contributing to high concentrations of intracellular drugs (Fig. 8)[90]. Ligand-targeted strategies may have particular potential to overcome drug resistance, especially those utilizing receptor-targeting ligands, since these ligands are typically internalized through endocytosis mediated by receptors. Indeed, doxorubicin[91] and transferrin-conjugated paclitaxel-loaded nanoparticles[92] containing folate receptor-targeted, pH-sensitive polymer micelle[92] inhibition was stronger for drug-resistant MCF-7 cells and/or xenografts than those for a non-target-free drug forms.

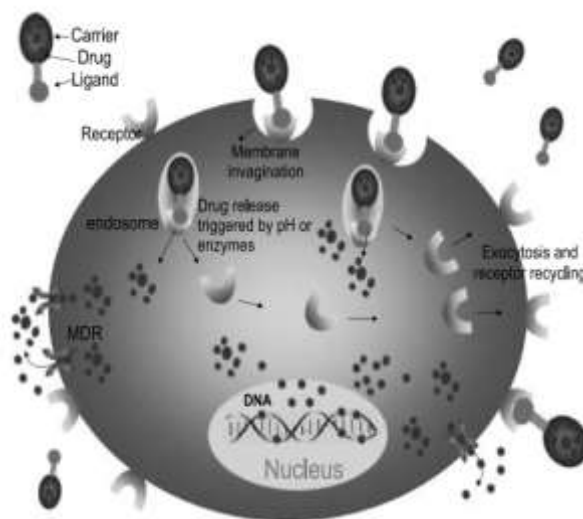


Fig.8 Internalization through nanoparticles to be internalized into the cell by the endosome. [92]

Targeted Drug Delivery Mechanism Transferrin Receptor

This approach to drug delivery was identified in a prostate cancer system. Compared to traditional paclitaxel distribution, Over a extended period of time, encapsulated paclitaxel nanoparticles transmit higher doses of paclitaxel into the cancer with surface-conjugated transferrin. The NP inhibits the use and related toxicity of Cremophor-EL vehicle-requiring paclitaxel by attacking cells of prostate cancer with upregulated receptors[93].



Electrochemical Nanosensor

In various sectors such as medicine, medicinal, agricultural, water, waste water, food contributing to health, protection and human property, electrochemistry has provided a strong and successful analytical method for scientific purposes. The most common instruments used commonly for the various studies are voltammetry, amperometry, and potentiometry. Electrochemical (bio) sensors translate electrochemical data to electrical signals that are detectable. The technologies most widely used in electrochemical sensors are cyclic voltammetry (CV), linear sweep voltammetry (LSV), anodic stripping voltammetry (ASV), and differential pulse voltammetry (DPV). The market for point-of-care applications is a significant driving force behind the production of nanoelectrochemical devices. Owing to their limited footprint and improved sensitivity capacity, nanoelectrode devices provide an appealing choice in this area. A number of medically important analytes, ranging from simple molecules such as histamine and dopamine to larger ones, were used to classify them. As a consequence of the versatility and range of nanoelectrode devices, more complicated objects such as bacteria or pharmaceutical drugs, because of the selectivity of enzymes as biorecognition components, enzyme dependent biosensors have received much interest. Combined with recent advances in nanoelectrochemistry, this selectivity has resulted in the advent of highly precise and sensitive biosensors. Composed of a single nanowire electrodes developed using a hybrid e-beam/optical lithography approach are often used to detect glucose at a depth of 10 μM using a mediated detection technique[94]. A typical technique for enhancing sensor efficiency with carbon nanotubes, nanorods, and gold nanoparticles is to decorate macro or microelectrodes with nanoparticles, Both of which find uses for the sensing of glucose. The presence of such nanoparticles increases the roughness of the surface area and larger electrodes, while electrocatalytic properties for electroactive compounds can also be beneficial. [95].

Conclusions

By manipulating a community of diseases, drug delivery is extremely delusional, and drug delivery methods have been established in current studies by connecting them with nanomaterials through the use of nanotechnology technology in the treatment of cancer and manipulating it, as is the case, by direct guidance to cancer cells without side effects. With chemotherapy in the sense of its side effects on carcinoma disease, as well as modern techniques to connect these techniques with biosensors to test their effects on the one hand and to monitor them to provide the highest selectivity and the best sensitivity to achieve separate results.

References

1. Siegel, R. L., Miller, K. D., and Jemal, A. (2020). Cancer statistics, 2020. *CA Cancer J. Clin.* 70, 7–30. doi: 10.3322/caac.21590
2. Zitvogel, L., Apetoh, L., Ghiringhelli, F., and Kroemer, G. (2008) Immunological aspects of cancer chemotherapy. *Nat. Rev. Immunol.* 8, 59–73. doi: 10.1038/nri2216
3. Dadwal, A., Baldi, A., and Kumar Narang, R. (2018). Nanoparticles as carriers for drug delivery in cancer. *Artif. Cells Nanomed. Biotechnol.* 46, 295–305. doi: 10.1080/21691401.2018.1457039



4. Palazzolo, S., Bayda, S., Hadla, M., Caligiuri, I., Corona, G., Toffoli, G., et al. (2018). The clinical translation of organic nanomaterials for cancer therapy: a focus on polymeric nanoparticles, micelles, liposomes and exosomes. *Curr. Med. Chem.* 25, 4224–4268. doi: 10.2174/0929867324666170830113755
5. E. Al-jawadi, M. Majeed. Electrochemical Sensors Based on Poly (L-Phenyl alanine) Film on MWCNT for Determination of TPS. *Periódico Tchê Química*.17(35): p 579-590. (2020).
6. Kipp, J. E. (2004). The role of solid nanoparticle technology in the parenteral delivery of poorly water-soluble drugs. *Int. J. Pharm.* 284, 109–122. doi: 10.1016/j.ijpharm.2004.07.019
7. Zhang, L., Chan, J. M., Gu, F. X., Rhee, J. W., Wang, A. Z., Radovic-Moreno, A. F., et al. (2008). Self-assembled lipid–polymer hybrid nanoparticles: a robust drug delivery platform. *ACS Nano* 2, 1696–1702. doi: 10.1021/nn800275r
8. Bertrand, N., Wu, J., Xu, X., Kamaly, N., and Farokhzad, O. C. (2014). Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. *Adv. Drug Deliv. Rev.* 66, 2–25. doi: 10.1016/j.addr.2013.11.009
9. Kalyane, D., Raval, N., Maheshwari, R., Tambe, V., Kalia, K., and Tekade, R. K. (2019). Employment of enhanced permeability and retention effect (EPR): nanoparticle-based precision tools for targeting of therapeutic and diagnostic agent in cancer. *Mater. Sci. Eng. C Mater. Biol. Appl.* 98, 1252–1276. doi: 10.1016/j.msec.2019.01.066
10. O'Brien, M. E., Wigler, N., Inbar, M., Rosso, R., Grischke, E., Santoro, A., et al. (2004). Reduced cardiotoxicity and comparable efficacy in a phase III trial of pegylated liposomal doxorubicin HCl (CAELYX/Doxil) versus conventional doxorubicin for first-line treatment of metastatic breast cancer. *Ann. Oncol.* 15, 440–449. doi: 10.1093/annonc/mdh097
11. Cortes, J., and Saura, C. (2010). Nanoparticle albumin-bound (nab (TM))paclitaxel: improving efficacy and tolerability by targeted drug delivery in metastatic breast cancer. *EJC Suppl.* 8, 1–10. doi: 10.1016/s1359-6349(10)70002-1
12. Riley, R. S., and Day, E. S. (2017). Gold nanoparticle-mediated photothermal therapy: applications and opportunities for multimodal cancer treatment. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* 9:e1449. doi: 10.1002/wnan.1449
13. Yoon, H. Y., Selvan, S. T., Yang, Y., Kim, M. J., Yi, D. K., Kwon, I. C., et al. (2018). Engineering nanoparticle strategies for effective cancer immunotherapy. *Biomaterials* 178, 597–607. doi: 10.1016/j.biomaterials.2018.03.036
14. Zang, X., Zhao, X., Hu, H., Qiao, M., Deng, Y., and Chen, D. (2017). Nanoparticles for tumor immunotherapy. *Eur. J. Pharm. Biopharm.* 115, 243–256. doi: 10.1016/j.ejpb.2017.03.013
15. Davis, M. E., Zuckerman, J. E., Choi, C. H., Seligson, D., Tolcher, A., Alabi, C. A., et al. (2010). Evidence of RNAi in humans from systemically administered siRNA via targeted nanoparticles. *Nature* 464, 1067–1070. doi: 10.1038/nature08956
16. Hrkach, J., Von Hoff, D., Mukkaram Ali, M., Andrianova, E., Auer, J., Campbell, T., et al. (2012). Preclinical development and clinical translation of a PSMAtargeted docetaxel nanoparticle with a differentiated pharmacological profile. *Sci. Transl. Med.* 4:128ra139. doi:



- 10.1126/scitranslmed.3003651
17. Mottaghitalab, F., Farokhi, M., Fatahi, Y., Atyabi, F., and Dinarvand, R. (2019). New insights into designing hybrid nanoparticles for lung cancer: diagnosis and treatment. *J. Control Release* 295, 250–267. doi: 10.1016/j.jconrel.2019.01.009
 18. Li, W., Zhang, H., Assaraf, Y. G., Zhao, K., Xu, X., Xie, J., et al. (2016). Overcoming ABC transporter-mediated multidrug resistance: molecular mechanisms and novel therapeutic drug strategies. *Drug Resist. Updat.* 27, 14–29. doi: 10.1016/j.drug.2016.05.001
 19. Alimoradi, H., Greish, K., Barzegar-Fallah, A., Alshaibani, L., and Pittalà, V. (2018). Nitric oxide-releasing nanoparticles improve doxorubicin anticancer activity. *Int. J. Nanomed.* 13, 7771–7787. doi: 10.2147/ij.n.s187089
 20. Wang, H., Agarwal, P., Zhao, G., Ji, G., Jewell, C. M., Fisher, J. P., et al. (2018b). Overcoming Ovarian Cancer Drug Resistance with a Cold Responsive Nanomaterial. *ACS Cent. Sci.* 4, 567–581.
 21. Zhang, J., Wang, L., You, X., Xian, T., Wu, J., and Pang, J. (2019). Nanoparticle therapy for prostate cancer: overview and perspectives. *Curr. Top. Med. Chem.* 19, 57–73. doi: 10.2174/1568026619666190125145836
 22. Hughes GA. Nanostructure-mediated drug delivery. *Nanomed Nanotech Biol Med* 2005;1:22–30.
 23. Moghimi SM, Hunter AC, Murray JC. Nanomedicine: Current status and future prospects. *FASEB J* 2005;19:311–30.
 24. F. Tajarobi, M. El-Sayed, B. D. Rege, J. E. Polli, and H. Ghandehari, “Transport of poly amidoamine dendrimers across Madin-Darby canine kidney cells,” *International Journal of Pharmaceutics*, vol. 215, no. 1-2, pp. 263–267, 2001.
 25. J. Frechet, “Functional polymers and dendrimers: reactivity, molecular architecture, and interfacial energy,” *Science*, vol. 263, no. 5154, pp. 1710–1715, 1994.
 26. M. El-Sayed, M. Ginski, C. Rhodes, and H. Ghandehari, “Transepithelial transport of poly(amidoamine) dendrimers across Caco-2 cell monolayers,” *Journal of Controlled Release*, vol. 81, no. 3, pp. 355–365, 2002.
 27. K. M. Kitchens, R. B. Kolhatkar, P. W. Swaan, N. D. Eddington, and H. Ghandehari, “Transport of poly(amidoamine) dendrimers across Caco-2 cell monolayers: influence of size, charge and fluorescent labeling,” *Pharmaceutical Research*, vol. 23, no. 12, pp. 2818–2826, 2006.
 28. R. B. Kolhatkar, P. Swaan, and H. Ghandehari, “Potential oral delivery of 7-ethyl-10-hydroxycamptothecin (SN-38) using poly(amidoamine) dendrimers,” *Pharmaceutical Research*, vol. 25, no. 7, pp. 1723–1729, 2008.
 29. M. Najlah, S. Freeman, D. Attwood, and A. D’Emanuele, “In vitro evaluation of dendrimer prodrugs for oral drug delivery,” *International Journal of Pharmaceutics*, vol. 336, no. 1, pp. 183–190, 2007.
 30. N. Vijayalakshmi, A. Ray, A. Malugin, and H. Ghandehari, “Carboxyl-terminated PAMAM-SN38 conjugates: synthesis, characterization, and in vitro evaluation,” *Bioconjugate Chemistry*, vol. 21, no. 10, pp. 1804–1810, 2010.
 31. D. A. Tomalia, “Starburst/cascade dendrimers: fundamental building blocks for a new nanoscopic chemistry set,” *Advanced Materials*, vol. 6, no. 7-8, pp. 529–539, 1994.



32. D. S. Wilbur, P. M. Pathare, D. K. Hamlin, K. R. Buhler, and R. L. Vessella, "Biotin reagents for antibody pretargeting. 3. Synthesis, radioiodination, and evaluation of biotinylated starburst dendrimers," *Bioconjugate Chemistry*, vol. 9, no. 6, pp. 813–825, 1998.
33. N. A. Peppas, "Star polymers and dendrimers: prospects of their use in drug delivery and pharmaceutical applications," *Controlled Release Society Newsletter*, vol. 12, pp. 12-13, 1995.
34. R. Jevprasesphant, J. Penny, D. Attwood, and A. D'Emanuele, "Transport of dendrimer nanocarriers through epithelial cells via the transcellular route," *Journal of Controlled Release*, vol. 97, no. 2, pp. 259–267, 2004.
35. M. El-Sayed, C. A. Rhodes, M. Ginski, and H. Ghandehari, "Transport mechanism(s) of poly (amidoamine) dendrimers across Caco-2 cell monolayers," *International Journal of Pharmaceutics*, vol. 265, no. 1-2, pp. 151–157, 2003.
36. T. Florence, T. Sakthivel, and I. Toth, "Oral uptake and translocation of a polylysine dendrimer with a lipid surface," *Journal of Controlled Release*, vol. 65, no. 1-2, pp. 253–259, 2000.
37. R. Wiwattanapatapee, B. Carreno-Gomez, N. Malik, and R. Duncan, "PAMAM dendrimers as a potential oral drug delivery system: uptake by everted rat intestinal sacs in-vitro," *Journal of Pharmacy and Pharmacology*, vol. 50, no. S9, p. 99, 1998.
38. N. Malik, R. Wiwattanapatapee, R. Klopsch et al., "Dendrimers: relationship between structure and biocompatibility in vitro, and preliminary studies on the biodistribution of ¹²⁵I-labelled polyamidoamine dendrimers in vivo," *Journal of Controlled Release*, vol. 65, no. 1-2, pp. 133–148, 2000.
39. R. Wiwattanapatapee, B. Carreno-Gomez, N. Malik, and R. Duncan, "Anionic PAMAM dendrimers rapidly cross adult rat intestine in vitro: a potential oral delivery system?," *Pharmaceutical Research*, vol. 17, no. 8, pp. 991–998, 2000.
40. D. Mullin, "Prometheus in Gloucestershire: Edward Jenner, 1749–1823," *The Journal of Allergy and Clinical Immunology*, vol. 112, no. 4, pp. 810–814, 2003.
41. Y. Lvov, K. Ariga, I. Ichinose, and T. Kunitake, "Assembly of multicomponent protein films by means of electrostatic layer-by-layer adsorption," *Journal of the American Chemical Society*, vol. 117, no. 22, pp. 6117–6123, 1995.
42. Y. M. Lvov, Z. Lu, J. B. Schenkman, X. Zu, and J. F. Rusling, "Direct electrochemistry of myoglobin and cytochrome P450 in alternate layer-by-layer films with DNA and other polyions," *Journal of the American Chemical Society*, vol. 120, no. 17, pp. 4073–4080, 1998.
43. G. Decher, B. Lehr, K. Lowack, Y. Lvov, and J. Schmitt, "New nanocomposite films for biosensors: layer-by-layer adsorbed films of polyelectrolytes, proteins or DNA," *Biosensors and Bioelectronics*, vol. 9, no. 9-10, pp. 677–684, 1994.
44. Y. Lvov, G. Decher, and G. Sukhorukov, "Assembly of thin films by means of successive deposition of alternate layers of DNA and poly(allylamine)," *Macromolecules*, vol. 26, no. 20, pp. 5396–5399, 1993.
45. Y. Lvov, H. Haas, G. Decher et al., "Successive deposition of alternate layers of polyelectrolytes and a charged virus," *Langmuir*, vol. 10, no. 11, pp. 4232–4236, 1994.



46. S. Watanabe and S. L. Regen, "Dendrimers as building blocks for multilayer construction," *Journal of the American Chemical Society*, vol. 116, no. 19, pp. 8855-8856, 1994.
47. P. A. Fiorito, V. R. Gonçalves, E. A. Ponzio, and S. I. C. de Torresi, "Synthesis, characterization and immobilization of Prussian blue nanoparticles. A potential tool for biosensing devices," *Chemical Communications*, vol. 21, no. 3, pp. 366-368, 2005.
48. F. Patolsky, T. Gabriel, and I. Willner, "Controlled electrocatalysis by microperoxidase-11 and Au-nanoparticle superstructures on conductive supports," *Journal of Electroanalytical Chemistry*, vol. 479, no. 1, pp. 69-73, 1999.
49. E. Al-Jawadi, M. Majeed. Detection of irinotecan using titanium nanoparticules modified electrode. *Bull. Chem. Soc. Ethiop.* **34**(2):p. 227-236.(2020). DOI: <https://dx.doi.org/10.4314/bcse.v34i2.2>
50. P. G. Pickup, W. Kutner, C. R. Leidner, and R. W. Murray, "Redox conduction in single and bilayer films of redox polymer," *Journal of the American Chemical Society*, vol. 106, no. 7, pp. 1991-1998, 1984.
51. G. P. Kittlesen, H. S. White, and M. S. Wrighton, "Chemical derivatization of microelectrode arrays by oxidation of pyrrole and N-methylpyrrole: fabrication of molecule-based electronic devices," *Journal of the American Chemical Society*, vol. 106, no. 24, pp. 7389-7396, 1984.
52. Rubinstein, "Electrostatic encapsulation using bi-polar polymer films on electrodes," *Journal of Electroanalytical Chemistry*, vol. 195, no. 2, pp. 431-434, 1985.
53. Rubinstein and I. Rubinstein, "Bipolar polymeric arrangements on electrodes," *Journal of Physical Chemistry*, vol. 91, no. 1, pp. 235-241, 1987.
54. D. A. Tomalia and J. M. J. Fréchet, "Discovery of dendrimers and dendritic polymers: a brief historical perspective," *Journal of Polymer Science Part A: Polymer Chemistry*, vol. 40, no. 16, pp. 2719-2728, 2002.
55. V. V. Tsukruk, "Assembly of supramolecular polymers in ultrathin films," *Progress in Polymer Science*, vol. 22, no. 2, pp. 247-311, 1997.
56. V. V. Tsukruk and J. H. Wendorff, "Supramolecular polymers and assemblies: mesomorphism and beyond," *Trends in Polymer Science*, vol. 3, no. 3, pp. 82-89, 1995.
57. D. A. Tomalia, A. M. Naylor, and W. A. Goddard, "Starburst dendrimers: molecular-level control of size, shape, surface chemistry, topology, and flexibility from atoms to macroscopic matter," *Angewandte Chemie International Edition*, vol. 29, no. 2, pp. 138-175, 1990.
58. J. Ledesma-García, J. Manríquez, S. Gutiérrez-Granados, and L. A. Godínez, "Dendrimer modified thiolated gold surfaces as sensor devices for halogenated alkyl-carboxylic acids in aqueous medium. A promising new type of surfaces for electroanalytical applications," *Electroanalysis*, vol. 15, no. 7, pp. 659-666, 2003.
59. L. Shen and N. Hu, "Heme protein films with polyamidoamine dendrimer: direct electrochemistry and electrocatalysis," *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, vol. 1608, no. 1, pp. 23-33, 2004.
60. H. C. Yoon and H.-S. Kim, "Multilayered assembly of dendrimers with enzymes on gold: thickness-



- controlled biosensing interface,” *Analytical Chemistry*, vol. 72, no. 5, pp. 922–926, 2000.
61. B. Alonso, P. Garcia Armada, J. Losada, I. Cuadrado, B. Gonzalez, and C. M. Casado, “Amperometric enzyme electrodes for aerobic and anaerobic glucose monitoring prepared by glucose oxidase immobilized in mixed ferrocene-cobaltocenium dendrimers,” *Biosensors and Bioelectronics*, vol. 19, no. 12, pp. 1617–1625, 2004.
 62. Brigger I, Dubernet C, Couvreur P. Nanoparticles in cancer therapy and diagnosis. *Adv Drug Deliv Rev* 2002;54:631–51.
 63. Gulati M, Grover M, Singh S and Singh M: Lipophilic drug derivatives in liposomes. *Int J Pharm* 165: 129-168, 1998.
 64. Scherphof G, Roerdink F, Waite M and Parks J: Disintegration of phosphatidylcholine liposomes in plasma as a result of interaction with high-density lipoproteins. *Biochim Biophys Acta* 542:296-307, 1978.
 65. Allen TM and Cleland LG: Serum-induced leakage of liposome contents. *Biochim Biophys Acta* 597: 418-426, 1980.
 66. Senior J and Gregoriadis G: Is half-life of circulating liposomes determined by changes in their permeability? *FEBS Lett* 145: 109-114, 1982.
 67. Yu L, Dean K and Li L: Polymer blends and composites from renewable resources. *Prog Polym Sci* 31: 576-602, 2006.
 68. Jain RK and Stylianopoulos T: Delivering nanomedicine to solid tumors. *Nat Rev Clin Oncol* 7: 653-664, 2010.
 69. Kamaly N, Yameen B, Wu J and Farokhzad OC: Degradable controlled-release polymers and polymeric nanoparticles: Mechanisms of controlling drug release. *Chem Rev* 116: 2602-2663, 2016.
 70. Holback H and Yeo Y: Intratumoral drug delivery with nanoparticulate carriers. *Pharm Res* 28: 1819-1830, 2011.
 71. Jain RK, Martin JD and Stylianopoulos T: The role of mechanical forces in tumor growth and therapy. *Annu Rev Biomed Eng* 16: 321-346, 2014.
 72. Kim KY: Nanotechnology platforms and physiological challenges for cancer therapeutics. *Nanomedicine (Lond)* 3: 103-110, 2007.
 73. Bahrami, B., Hojjat-Farsangi, M., Mohammadi, H., Anvari, E., Ghalamfarsa, G., Yousefi, M., et al. (2017). Nanoparticles and targeted drug delivery in cancer therapy. *Immunol. Lett.* 190, 64–83. doi: 10.1016/j.imlet.2017.07.015
 74. Venturoli, D., and Rippe, B. (2005). Ficoll and dextran vs. globular proteins as probes for testing glomerular permselectivity: effects of molecular size, shape, charge, and deformability. *Am. J. Physiol. Renal. Physiol.* 288, F605–F613. doi:10.1152/ajprenal.00171.2004
 75. Decuzzi, P., Pasqualini, R., Arap, W., and Ferrari, M. (2009). Intravascular delivery of particulate systems: does geometry really matter? *Pharm. Res.* 26, 235–243. doi: 10.1007/s11095-008-9697-x
 76. Yang, Q., Jones, S. W., Parker, C. L., Zamboni, W. C., Bear, J. E., and Lai, S. K. (2014). Evading immune cell uptake and clearance requires PEG grafting at densities substantially exceeding the



- minimum for brush conformation. *Mol. Pharm.* 11, 1250–1258. doi: 10.1021/mp400703d
77. Perrault, S. D., Walkey, C., Jennings, T., Fischer, H. C., and Chan, W. C. (2009). Mediating tumor targeting efficiency of nanoparticles through design. *Nano Lett.* 9, 1909–1915. doi: 10.1021/nl900031y
 78. Wong, C., Stylianopoulos, T., Cui, J., Martin, J., Chauhan, V. P., Jiang, W., et al. (2011). Multistage nanoparticle delivery system for deep penetration into tumor tissue. *Proc. Natl. Acad. Sci. U.S.A.* 108, 2426–2431. doi: 10.1073/pnas.1018382108
 79. Amreddy, N., Muralidharan, R., Babu, A., Mehta, M., Johnson, E. V., Zhao, Y. D., et al. (2015). Tumor-targeted and pH-controlled delivery of doxorubicin using gold nanorods for lung cancer therapy. *Int. J. Nanomed.* 10, 6773–6788. doi:10.2147/ijn.s93237
 80. Liu, L., Wei, Y., Zhai, S., Chen, Q., and Xing, D. (2015). Dihydroartemisinin and transferrin dual-dressed nano-graphene oxide for a pH-triggered chemotherapy. *Biomaterials* 62, 35–46. doi: 10.1016/j.biomaterials.2015.05.036
 81. Santi, M., Maccari, G., Mereghetti, P., Voliani, V., Rocchiccioli, S., Ucciferri, N., et al. (2017). Rational Design of a Transferrin-Binding Peptide Sequence Tailored to Targeted Nanoparticle Internalization. *Bioconjug. Chem.* 28, 471–480. doi: 10.1021/acs.bioconjchem.6b00611
 82. Cui, Y. N., Xu, Q. X., Davoodi, P., Wang, D. P., and Wang, C. H. (2017). Enhanced intracellular delivery and controlled drug release of magnetic PLGA nanoparticles modified with transferrin. *Acta Pharmacol. Sin.* 38, 943–953. doi: 10.1038/aps.2017.45
 83. Soe, Z. C., Kwon, J. B., Thapa, R. K., Ou, W., Nguyen, H. T., Gautam, M., et al. (2019). Transferrin-conjugated polymeric nanoparticle for receptor-mediated delivery of doxorubicin in Doxorubicin-resistant breast cancer cells. *Pharmaceutics* 11:63. doi: 10.3390/pharmaceutics11020063
 84. Litman, T., Brangi, M., Hudson, E., Fetsch, P., Abati, A., Ross, D. D., et al. (2000). The multidrug-resistant phenotype associated with overexpression of the new ABC half-transporter. MXR (ABCG2). *J. Cell Sci.* 113(Pt 11), 2011–2021.
 85. Moghimi SM, Hunter AC, Murray JC. Long-circulating and target-specific nanoparticles: theory to practice. *Pharmacol Rev* 2001;53:283^ 318.
 86. Wisse E, Braet F, Luo D, et al. Structure and function of sinusoidal lining cells in the liver. *Toxicol Pathol* 1996; 24 : 10 0 ^ 11.
 87. Yuan F, Dellian M, Fukumura D, et al. Vascular permeability in a human tumor xenograft: molecular size dependence and cutoff size. *Cancer Res* 1995;55: 3752^ 6.
 88. Moghimi SM, Szebeni J. Stealth liposomes and long circulating nanoparticles: critical issues in pharmacokinetics, opsonization and protein-binding properties. *Prog Lipid Res* 2003;42:463^ 78.
 89. Gottesman MM, FojoT, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev Cancer* 2002;2:48^ 58.
 90. Wong HL, Bendayan R, Rauth AM, Xue HY, Babakhanian K, Wu XY. A mechanistic study of enhanced doxorubicin uptake and retention in multidrug resistant breast cancer cells using a polymerlipid hybrid nanoparticle system. *J Pharmacol Exp Ther* 2006;317:1372^ 81.
 91. Lee ES, Na K, Bae YH. Doxorubicin loaded pHsensitive polymeric micelles for reversal of resistant



- MCF-7 tumor. *J Control Release* 2005;103: 405[^]18.
92. Sahoo SK, Labhasetwar V. Enhanced antiproliferative activity of transferrin-conjugated paclitaxel-loaded nanoparticles is mediated via sustained intracellular drug retention. *Mol Pharm* 2005;2:373[^] 83.
 93. Sahoo SK, Ma W, Labhasetwar V. Efficacy of transferrin-conjugated paclitaxel-loaded nanoparticles in a murine model of prostate cancer. *Int J Cancer* 2004;112:335– 40.
 94. Tian K, Alex S, Siegel G, Tiwari A. Enzymatic glucose sensor based on Au nanoparticle and plant-like ZnO film modified electrode. *Mater Sci Eng C Mater Biol Appl.* 2015;46:548–552.
 95. Wang HH, Bu Y, Dai WL, Li K, Wang HD, Zuo X. Well-dispersed cobalt phthalocyanine nanorods on graphene for the electrochemical detection of hydrogen peroxide and glucose sensing. *Sensors and Actuators B: Chemical.* 2015;216:298–306.