



**IMMUNOHISTOCHEMICAL ASSAY USED TO CORRELATE LYMPHOCYTES CD3, CD8  
IN PATIENTS WITH GASTRIC ADENOCARCINOMA**

Noor Al- Huda Ali A. H. Saeed

Biology Department, College of Science, Mustansiriyah University. Iraq.

Sara Ibraheem Mahmood

Biology Department, College of Science, Mustansiriyah University. Iraq.

Liqaa Jameel Ibraheem

Biology Department, College of Science, Mustansiriyah University. Iraq.

E. mail: nooral\_huda@uomustansiriyah.edu.iq

**Abstract**

This study's objectives were to define the relationship between lymphocytes CD3 and CD8 in patients with gastric adenocarcinoma and to identify these two markers in connection to some patient characteristics (patient age, gender, grade, and stage). By using an immunohistochemical assay, the expression of the relationship between CD3, CD8, and gastric cancer was evaluated. According to the findings, stomach tissue samples were taken from 60 patients who had gastrectomy and had gastric cancer. The collection of these samples took place between June 2021 and February 2022. Sixty patients included 40 men and 20 women, their ages ranged between (27-74) years, with mean age 56 years. Out of 60 patients, 34 (56.66%) had immunohistochemistry evidence of the lymphocytes marker CD3. Thus, out of 60 samples, the CD8 was found in 26 (43.3%). and the middle and lower thirds of the stomach were the only locations for the double malignancies in stages 0, I, and II, with the histological grade including 35, 21, and 4 (poor, well, and moderate) differentiation, respectively. Additionally, results showed that there was a considerably stronger correlation between CD3 positivity and each of age, gender, histological grade, and tumor stage. While there was no significant relationship between CD8 expression and the patients' ages, the positive results in cytotoxic T-lymphocyte CD8 were considerably greater associated to each of grade, gender, and histological stage of the tumor.

**Introduction**

The second most frequent cancer in the world is gastric cancer [1]. Stomach cancer, also known as gastric cancer, is a type of cancer that can develop in any area of the stomach and accounts for over 800,000 fatalities annually worldwide [2]. The most frequent type of stomach cancer is adenocarcinoma, which begins in the glandular tissue of the stomach and accounts for 90 to 95 percent of all stomach cancer. Stomach cancers are classified by the type of tissue where they develop [3]. According to the physical characteristics of the tumor, Lauren divided gastric adeno in two histological kinds, intestinal and diffuse [4,5]. Incidence and fatality rates for stomach cancer have generally decreased recently. Gastric carcinoma is still the most prevalent type of cancer in Asian nations [6, 7]. It frequently involves both inherent predisposition and environmental influences, making it complex.



Diet, *Helicobacter pylori* infection, prior stomach surgery, atrophic gastritis, and radiation exposure are environmental variables linked to the onset of gastric cancer[8,9]. Although the prognosis for patients with gastric cancer has not yet been thoroughly investigated, it is generally accepted that immune cell infiltration within a tumor and tumor progression are related. As a result, the distribution of different immune cell types found in gastric cancer may offer helpful information regarding the prognosis of the patient [6,7]. Important cellular elements of the adaptive immune system are T cells. They are triggered to serve as helpers, effectors, or regulators by particular antigens in order to mount and coordinate an effective immune response and to create immunological memory [10]. According to recent research [11,12], different kinds of tumor infiltrating lymphocytes (TIL) are linked to better clinical outcomes for a variety of human malignancies. The primary function of CD8 T-cells is to kill malignant or infected cells that are antigen-bearing [10]. In individuals with gastric carcinoma, the amount of CD8 T-cells found inside the cancer cell is a significant predictive factor[13]. The identifying of T-cells can be aided by CD3, which is a major indicator in the classification of aggressive tissue[14]. Peripheral blood lymphocytes (PBL) and tumor infiltrating lymphocytes (TIL) have both been found to have CD3 impairment[15]. CD3 antigen, previously believed to be unique to T-lymphocytes, has been discovered on gastric parietal cells in both humans and animals [16].

## Materials and Methods

Sixty Iraqi patients with stomach cancer provided paraffin embedded block samples. Someone have already had surgery at Baghdad Medical City between June 2021 and February 2022. Before the procedure, none of those patients had had any anticancer treatment. 40 men and 20 women, with a mean age of 56 and a range of 27 to 74 years, make up the male to female ratio of 2:1. Compared to 20 seemingly healthy controls, whose ages and sexual orientations were matched to the sick group. Immunohistochemistry was used to identify CD3 and CD8 in specimens, and the procedure was carried out as instructed in the kit's instruction leaflet.

-Immunohistochemistry served as the CD3 and CD8 detection method (IHC).

-Horseradish peroxidase, Streptavidin-Biotin 2 system, and universal Dakocytomation labeling (LSAB-2 system. HRP). code number Ko673, ready to use detection system (CA.USA). for (IHC).

-Polyclonal Rabbit Anti Human CD3 (A0452, Dakocytomation, Denmark), ready-to-use primary antibody.

-Primary antibody that is prepared for use (Monoclonal Mouse Anti Human CD8. Clone:C8/144B. Code No. Nr. M7103. Dako. Denmark.

## Immunohistochemical Assay

Using a microtome, sections of paraffin-embedded blocks were created that were 4 mm thick. These slices were all dehydrated and deparaffinized. These samples underwent a succession of dewaxing procedures in xylene (100, 90, 70, and D.W, respectively), after which they were put in an endogenous peroxidase block for 25 minutes. after adding CD3 or CD8 as the primary Ab. washing with PBS, secondary Ab was used and the samples were incubated for 1 hour, streptavidin in a humid chamber



for 30 minutes. counterstained using Mayer's hematoxyline, and serial ethanol (70,90,100%) and xylene dehydration.

## Statistical Analysis

The impact of different factors on research parameters was examined using the statistical analysis system, SAS [17]. Chi-square test at the Percentages comparison in this study.

## Results and Discussion

A total of 60 gastric adenocarcinoma specimens were examined immunohistochemically utilizing CD8 cytotoxic T-lymphocyte-detection monoclonal antibody and CD3 TIL polyclonal antibody. The findings revealed that the mean age of our patients was 56 years old, ranging from (27 to 74) years. Our findings were in line with a number of other papers [27,18,19] that stated the mean age was 56,58 and 57 years, respectively. 40 men and 20 women made up the male to female ratio in our study, which was 2:1. This discovery was supported by [20,27]. who stated that the ratio of men to women was 2:1, with 14 men and 8 women, and 32 men and 14 women, respectively. Our analysis found a highly significant and high percentage of tumors in stages I and II in both CD3 and CD8; [22] further supported this finding. They discovered that although the difference was not statistically significant, CD3 was lower in individuals with stage IV than those with stages I, II, and III. and [18]who discovered a strong relationship between CD8 and staging at ( $P=0.012$ ). Results also showed that poor differentiation cancer was found in 24 out of 46 patients, which is consistent with [21]. He said that stomach cancer was a highly cellular tumor made up of carcinoma cells that had not undergone proper differentiation. According to table 1, out of a total of 60 cases, positive CD3 results were found in 34 samples (56.66%)figure 1; this finding was supported by reports [23,24,27] showing CD3 expression in PBL from gastric cancer patients was considerably higher. Age, sex, grade, and the histological stage of the tumor all significantly correlated with CD3 TIL in gastric adenocarcinoma patients, respectively, (0.0086), (0.0096), ( 0.0225), and (0.0019). at ( $P < 0.01$ ). The results of the current investigation corroborated those of [25], who noted that stomach cancer patients had much greater CD3+ T-lymphocyte populations than healthy donors. According to table 2, 22 out of 60 cases (36.66%) had positive CD8 results figure 2, which contrasts with [27] who recorded (41.30%) of CD8 positive results and disagree with [18]'s claim that (96.9%) of cases had positive results. It was agreed with [26] who observed that CD8 ratio had no significant correlation with sex because results in table (2) indicated no significant correlation between CD8 and the age of the patients at ( $P= 0.0089$ ).figure3,4,5 and 6. As a result of this study's findings, it can be concluded that CD8 and CD3 have a highly significant relationship to the histological grade and stage of gastric cancer. This relationship was helpful in the clinical study of patients with gastric adenocarcinoma. Removal of the tumor may boost a patient's cellular immunity.



Table -1 shows the distribution of patients with gastric adenocarcinoma by age, sex, grade, and histological stage in relation to tumor infiltrating lymphocyte (TIL) CD3 levels.

The factor	CD3 positive% Total No.= 34 (56.66%)	CD3 Negative% Total No.=26 (43.33%)	p-value( $p \leq 0.01$ ) and $\chi^2$ value
Age			
≤ 40	12 (35.29%)	16 (61.53%)	0.0086
> 40	22 (64.70%)	10 (38.46%)	$\chi^2 = 9.362^{**}$
Grade			
poor	19 (55.88%)	14 (53.84%)	0.0225
Well	14 (41.17%)	9 (34.61%)	$\chi^2 = 7.456^{**}$
Moderate	1 (2.94%)	3 (11.53%)	
Gender			
Male	26 (76.47%)	14 (53.84%)	0.0096
Female	8 (23.58%)	12 (46.15%)	$\chi^2 = 9.240^{**}$
Stage			
0, I, II	22 (64.70%)	16 (61.53%)	0.0019
III, IV	12 (35.29%)	10 (38.46%)	$\chi^2 = 9.780^{**}$

$P < 0.01$  \*\* Highly significant

Table 2 : shows the distribution of patients with gastric adenocarcinoma by age, sex, grade, and histological stage in association to tumor infiltrating lymphocyte (TIL) CD8 levels.

The factor	CD8 positive% Total No.= 22 (36.66%)	CD8 Negative% Total No.= 38 (63.33%)	p- value ( $p \leq 0.01$ ) and $\chi^2$ value
Age			
≤ 40 28	9 (40.90%)	19 (50%)	0.0831
> 40 32	13 (59.09%)	19 (50%)	$\chi^2 = 2.654^* \text{ NS}$
Grade			
poor 35	13 (59.09%)	22 (57.89%)	0.0048
Well 21	8 (36.36%)	13 (34.21%)	$\chi^2 = 8.236^{**}$
Moderate 4	1 (4.54%)	3 (7.89%)	
Gender			
Male	14 (63.36%)	26 (68.42%)	0.0096
Female	8 (36.36%)	12 (31.57%)	$\chi^2 = 9.240^{**}$
Stage			
0, I, II	17 (72.27%)	21 (55.26%)	0.0089
III, IV	5(22.72%)	17 (44.73%)	$\chi^2 = 9.732^{**}$

$P < 0.01$

\*\* Highly significant

\*No significant

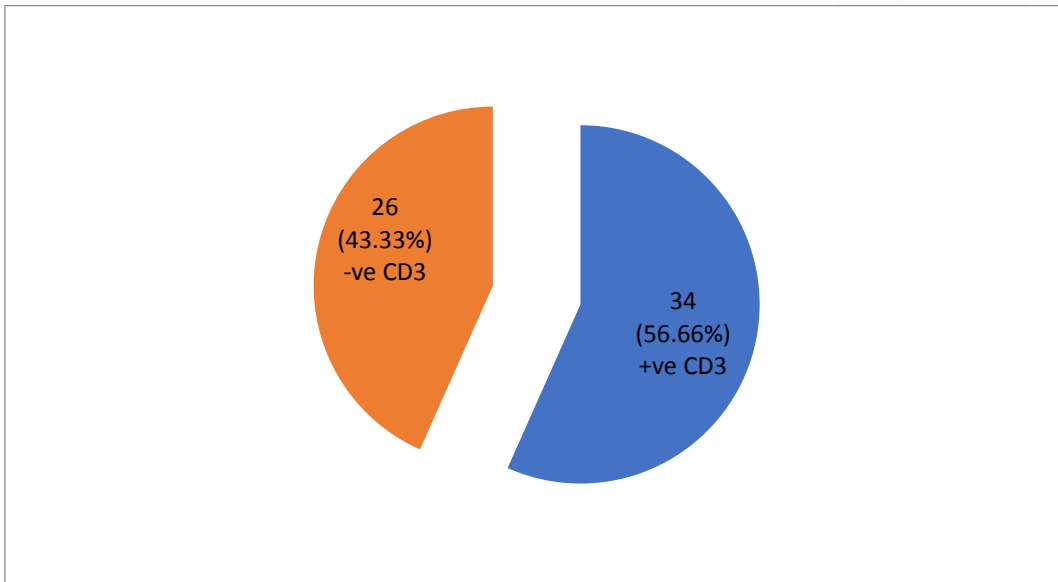


Figure 1: The percentage of adenocarcinomas with immunohistochemical reactivity for the immunological surface marker CD3.

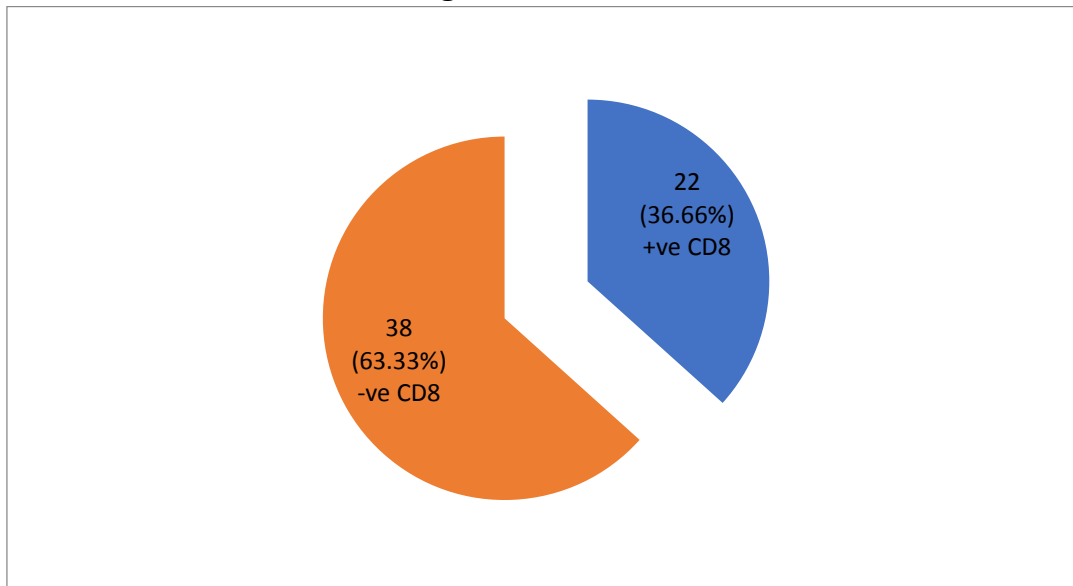


Figure 2: The percentage of adenocarcinomas with immunohistochemical reactivity for the immunological surface marker CD8.

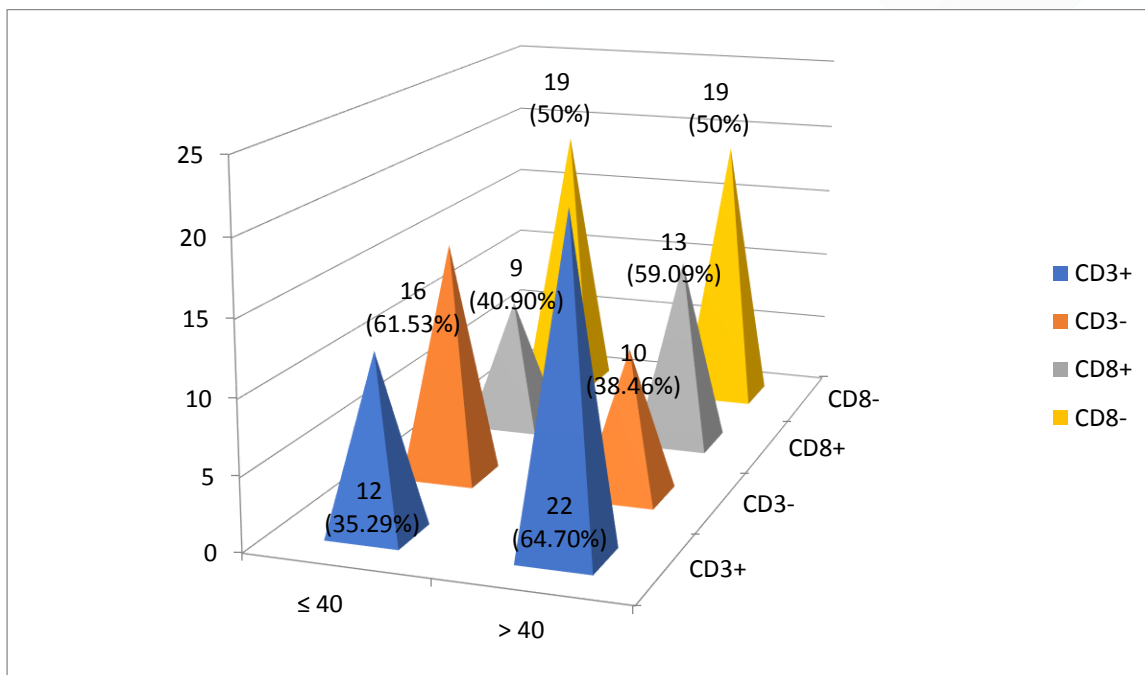


Figure 3: Distribution of adenocarcinoma patients according to their age in relation with CD3 and CD8 results

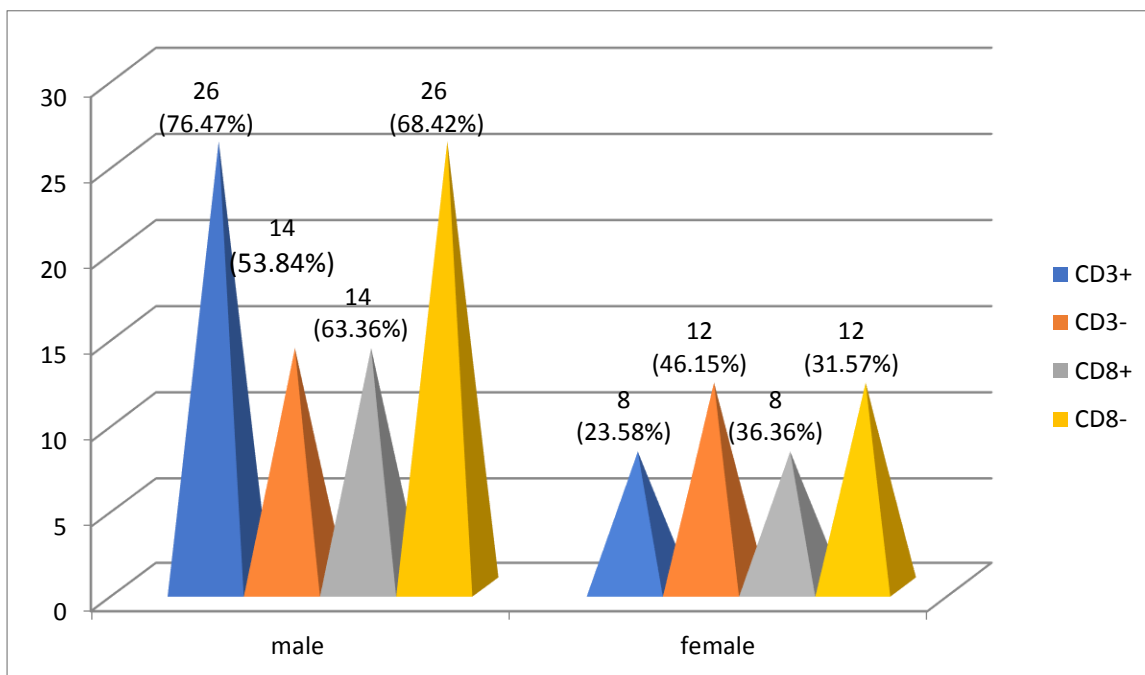


Figure 4: Distribution of adenocarcinoma patients according to their gender, in relation with CD3 and CD8 results

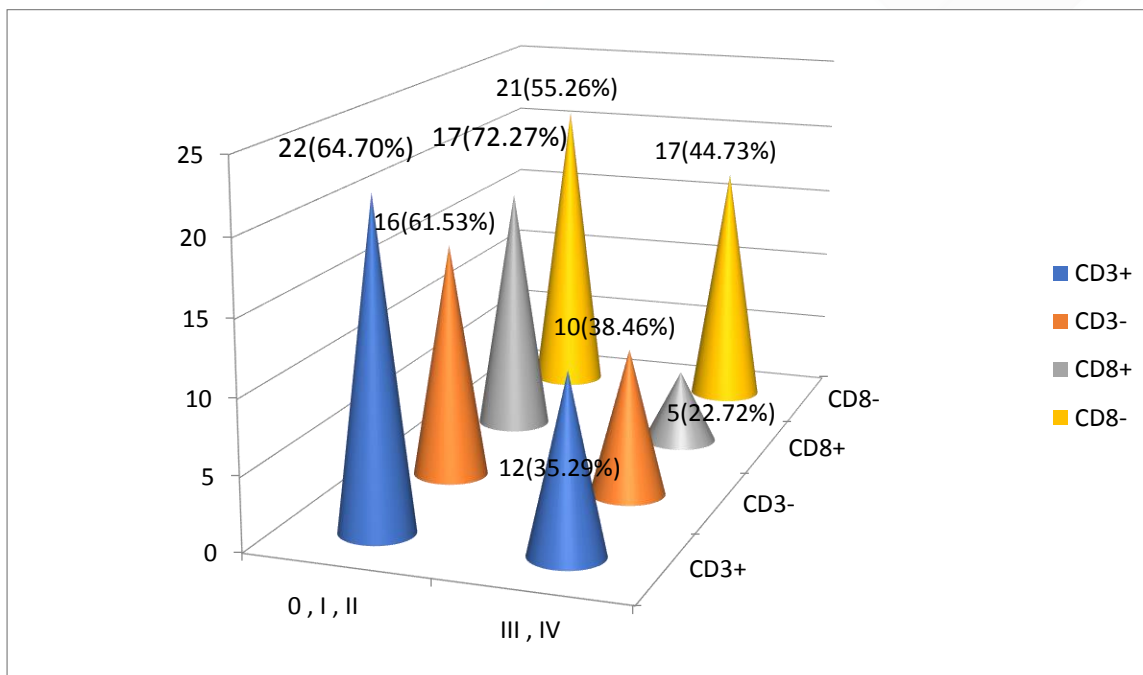


Figure 5: Distribution of adenocarcinoma patients according to the stage of the tumor, in relation with CD3 and CD8 results.

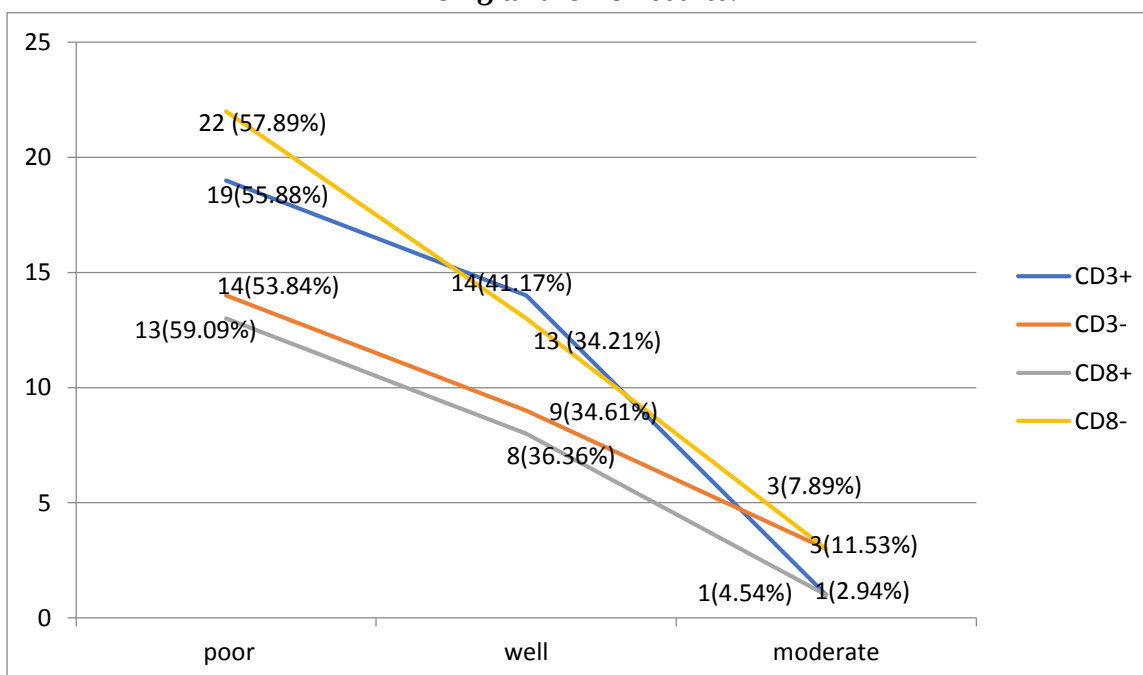


Figure 6: Distribution of adenocarcinoma patients according to the grade of the tumor, in relation with CD3 and CD8 results.





## Referances

- 1- Bozzetti F. , Marubini E., and Bonfanti G. , Subtotal versus total gastrectomy for gastric cancer : five- year survival rates in a multicenter randomized Italian trial. Italian Gastrointestinal Tumor Study Group. Ann Surg. 230 : 170-178,(1999).
- 2- 2- “ Cancer ( Fact sheet N° 297 )” World Health Organization. February (2009).
- 3- Gore R. , Gastrointestinal Cancer. Radiol Clin North Am. ; 35:295- 310, (1997).
- 4- Lauren P., The two histological main types of gastric carcinoma : diffuse and so-called ontestinal-type Carcinoma. Acta pathol . Microbiol. Scand., 64:31-49, (1965).
- 5- Correa P. , and Cheu V. W., Gastric cancer. In: R. Doll, J. F. Fraumeni, Jr. , and C. S. Muir (eds.), Trends in Cancer Incidence and Mortality, pp. 55-75. plainview, NY: Cold Spring Harbor Laboratory Press,(1994).
- 6- Jemal A., Siegel R., Ward E. etal. Cancer Statistics, CA Cancer J clin.; 57 (1):43-66,(2007).
- 7- Parkin D M., Bray F., Ferlay J. etal . Global Cancer Statistics, (2002).CA Cancer J Clin.; 55(2):74-108, (2005).
- 8- Bertuccio P. , Chatenoud L. , Levi F., etal ., “ Recent patterns in gastric cancer : a global overview” , International Journal of Cancer, vol. 125, no. 3, pp.666-673, (2009).
- 9- Chen J., Bu X. L. , Wang Q., Hu P. J. , and Chen M. H., “ Decreasing seroprevalence of helicobacter pylori infection during 1993-2003 in Guangzhou, Southern China” Helicobacter, vol. 12, no. 2, pp.164-169,(2007).
- 10- Castellino F., Germain R. N., Cooperation between CD4+ and CD8+ T cells : when, where, and how. Annu. Rev. Immunol. 24, 519-540,(2006).
- 11- Clemente CG., Mihm MC. Jr., Bufalino R., Zurrida S., Collini P., Cascinelli N., Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. Cancer; 77:1303-1310,(1996).
- 12- Taylor RC., Patel A., Panageas KS., Busam KJ., Brady MS., Tumor infiltrating lymphocytes predict sentinel lymph node positivity in patients with cutaneous melanoma. J Clin Oncol; 25:869-875,(2007).
- 13- Ohno S., Tachibana M., Fujii T., Ueda S., Kubota H., and Negasue N., Role of stromal collagen in immunodulation and prognosis of advanced gastric carcinoma . Int J Cancer 97:770-774,(2002).
- 14- Chetty R., Gatter K., CD3: Structure , Function, and role of immunostaining in clinical practice. J Pathol., 173 (4):1 , (1994).
- 15- De Gruijl TD., Bontkes HJ., Peccatori F., Gallee MP., Helmerhorst TJ., Verheijen RH., Aarbiou J., Mulder WM., Walboomers JM., Meijer CJ., Van de Vange N. and Scheper RJ., Expression of CD3 zeta on T-cells in primary cervical carcinoma and in metastasis positive and negative pelvic lymph nodes . Br J Cancer 79 : 1127- 32, (1999).
- 16- Banner B., Spicer Z., Alroy J., Expression of CD3 epsilon subunit in gastric parietal cells : a possible role in signal transduction. Pathol Res Pract.; 199(3): 137-43(2003).
- 17- SAS. Statistical Analysis System , User’s Guide. Statistical. Version 7th ed. SAS . Inst. Inc. Cary. N. C. USA.(2004).





- 18- Ju- gao C., Jian C. X., Xiao-ting L., Ke Pan, W. W., Lin L., Jing – Jing Z., Qi-J. W. , Yong qiany L., Shi ping C., Jia H., Li X.H., Miao-l. , Yi-bing C., Hai-qing M., Zhen- W. Z., Zhi-wei Z., Alfred E C., Qiao L., Intratumoral Expression of IL-7 and Its Prognostic Role in Gastric Adenocarcinoma Patients. *Int J Biol Sci* ; 7(1):53- 60,(2011).
- 19- Lee HE., Chae SW., Lee YJ., Kim MA., Lee HS., Lee BL. And Kim WH. , Prognostic implications of type and density of tumor infiltrating lymphocytes in gastric cancer. *Br J Cancer*. 99 (10 ):1704-1711(2008).
- 20- Vida M. , Grapina P., Violeta N., Renata J., Eugenijus S., Janina D., T- lymphocytes subset and lipid peroxidation in relation to survival among advanced gastric cancer patients. *Acta Medica Lituanica*. Vol.11 , no. 4, p. 19-23 (2004).
- 21- Toshinari M., Kishichiro W., Masayoshi M., Akishi O., Tokuji K., Yutaka T., Hiroshi U., Tomomi O. and Isao N., Clinico pathologic study of 27 cases and Immunohistochemical Analysis of the Subpopulations of infiltrating lymphocytes in the tumor. *Medullary Carcinoma with Lymphocytic Infiltration of the Stomach*. Vol.66 no. 5 p. 948(1990).
- 22- Cancer Reserch Center. Phenotypes and Clinical Significance of CD3 bright T-lymphocytes in Patients with gastric cancer.(2012).
- 23- Sumiya I., Shoji N., Futoshi M., Koki T., Akihiro N., Masataka M., Hiroshi O., Sabura N., Shuichi H., Ikuro M. and Takashi A.. CD3 Expression of Regional Lymph Node and Peripheral Blood Lymphocytes in Gastric Cancer. *Anti Cancer Reserch* 24:2123- 2126(2004).
- 24- Kim CW., Choi SH., Chung EJ., Lee MJ., Byun EK., Ryu MH. And Bang YJ:Alteration of signal transducing molecules and phenotypical characteristics in peripheral blood lymphocytes from gastric carcinoma patients. *Patho biology* 67:123-128(1999).
- 25- Osada J., Kamocki Z., Rusak M., Dabrowska M., Kedra B., The effect of surgical and nutritional treatment on activation parameters of peripheral blood T-lymphocytes in stomach cancer patients in postoperative period. *Pol Merkur Lekarski*; 24 (141):231-6(2008).
- 26- Lee WJ., Chang KJ., Lee CS., Chen KM., Selective depression of T-lymphocyte subsets in gastric cancer patients : an implication of immunotherapy. *J Surg Oncol*.:55 (3):165-9(1994).
- 27- Saeed, Noor Al-Huda Ali A. H.: Correlation Between Lymphocytes CD3,CD8 In Patients With Gastric Adenocarcinoma By Using Immunohistochemical Assay. *Al- Mustansiriyah J. Sci*. Vol. 24, No 2, (2013).